




Ozonated water electrolytically generated by diamond-coated electrodes controlled phytonematodes in replanted soil

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Abstract

Phytonematodes cause severe yield losses in horticulture, partly because they are difficult to manage. Compact, energy-efficient generators that electrochemically produce ozonated water by utilizing diamond-coated electrodes have become available. In this study, the application of on-site generated ozonated water to inactivate soil nematodes and to mitigate nematode-mediated apple replant disease was tested. *Pratylenchus penetrans* was highly susceptible to dissolved ozone (LC_{50} 0.6 mg L⁻¹). In one greenhouse experiment, treatment of *P. penetrans* in soil with ozonated water (0.27 mg ozone L⁻¹ soil) reduced subsequent invasion of the nematodes into roots by 60%. Growth of apple saplings in soil that was affected by apple replant disease (ARD) was significantly improved following a treatment with 1/10 volume ozonated water compared to the control. In a second greenhouse experiment, one-time drenching of ARD soil with ozonated water was followed by improved growth of apple plants similar to that in autoclaved soil. A second application of ozonated water did not further improve plant growth. The number of active nematodes in replanted soil that moved through a Baermann filter was significantly reduced by all tested concentrations of ozone (0.12–0.75 mg L⁻¹ soil). A fraction of 19–36% of the nematodes survived and slightly recovered after four weeks. In conclusion, on-site generated ozonated water has potential to mitigate nematode problems in horticulture and to expand management options.

Keywords Ozone treatment · Apple replant disease · Nematode · Pest control · Horticulture · Soil disinfestation

Introduction

Nematodes are abundant in soils and play an essential role in ecosystem functions and services (Yeates et al. 1993). Some of the nematodes, mainly the endoparasitic phytonematodes (Elling 2013; Jones et al. 2013), are a major limiting biotic factor of productivity (Singh et al. 2015; Stirling 2014). These nematodes migrate through soil in search of a host plant, invade roots and feed on the cytoplasm of cells, thereby damaging the plant. The endoparasitic root-lesion nematode *Pratylenchus penetrans* has a wide host range. In soybean, it can impede symbiotic nitrogen fixation and

plant growth (Elhady et al. 2020). Etiological relations between some nematodes and microbes in soil-borne disease complexes are known (Back et al. 2002; Morris et al. 2016). For example, *P. penetrans* was reported to exacerbate apple replant disease (ARD), in addition to unknown biotic factors (Dullahide et al. 1994; Jaffee et al. 1982; Oostenbrink and Hoestra 1961). ARD is characterized by reduced vegetative growth of apple trees, root damage and yield decline in replanted soil (Winkelmann et al. 2019). Soil disinfestation techniques directed towards nematodes such as heat at 50 °C (Yim et al. 2015), *Tagetes* as pre-plant (Yim et al. 2017), or application of nematicides (Jaffee et al. 1982; Johnson et al. 1982) resulted in improved plant growth. The peculiar role of nematodes in the development of ARD was further demonstrated by inoculation with nematodes or microbes on apple rootstocks, singly or in combination. ARD was induced in the presence of nematodes from replanted soil, while the microbes alone did not induce ARD but enhanced the disease together with the nematodes (Kanfra et al. 2018). Effective management of nematode-mediated diseases is still challenging.

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Application of synthetic nematicides poses health and environmental risks because of animal and human toxicity (Onkendi et al. 2014; Zasada et al. 2010). Crop rotation is of limited efficacy for plant-parasitic nematodes with a wide host range, and it is often not applicable in fruit and nursery production systems. Anaerobic soil disinfection or biofumigation with plants or seed meal of Brassicaceae could ameliorate ARD (Mazzola et al. 2015). However, these methods are limited by high time requirements, costs, logistical challenges, and the environmentally undesirable use of plastic. Nematode-related problems are further aggravated by the lack of resistant varieties or non-host crops (Cook and Starr 2006). Biological control options may be costly and of unpredictable efficacy (Agbenin 2012; Fourie et al. 2014). Hence, there is a need to exploit alternative mitigation strategies to protect food crops against nematode pests.

Ozone is a versatile and powerful disinfectant (Gray 2014). Dissolved ozone inactivates a wide range of microorganisms, insects, and nematodes (Al-Ahmadi et al. 2009; Perry and Yousef 2011; Qiu et al. 2009). The antimicrobial activity of ozone is attributed to its high oxidation potential. In addition, hydroxyl radicals arise from ozone in water and can break down cell walls and membranes. Ozone is generally regarded as safe because it has a unique property of auto-decomposition and leaves no toxic residues (Carletti et al. 2013). In contrast to gaseous ozone, dissolved ozone decomposes faster and does not pose a toxic risk to respiratory tracts (Kim et al. 2002) or plants (Fujiwara and Fujii 2002). Therefore, recent interest developed in using ozonated water for control of plant diseases (Fujiwara and Fujii 2002; Guo et al. 2019; Zheng et al. 2020).

Application of ozonated water to reduce plant-parasitic nematodes in soil was not comprehensively studied. This may be because ozone is expected to be inefficient in soil. It is assumed that a significant part of its oxidizing power is expended by the reaction with soil organic matter. Production of ozonated water had been highly inefficient until boron-doped diamond electrodes were developed for the continuous electrochemical generation of dissolved ozone directly from water (Arihara et al. 2007; Cobb et al. 2018). These electrodes became commercially available at various scales. As they are robust, energy-efficient and suitable for mobile devices, applications in agriculture and horticulture are becoming possible.

In this study, the hypothesis was tested whether ozonated water reduces nematode populations in soil and improves plant growth. This was tested by exposing *P. penetrans* to various concentrations of dissolved ozone. Concentration-dependent and frequency of application effects of ozonated water on inactivation of nematodes in ARD soil and mitigation of the disease were examined.

Materials and method

Electrochemical reactor systems for preparation of ozonated water

Ozonated water was generated using an electrochemical ozone generator (Condiapure™, Condias GmbH, Itzehoe, Germany) based on electrodes coated with boron-doped diamond that operate efficiently at low voltages. Ozone generation was controlled by operating the electrodes at a constant current (1.0–4.0 A) and 8–15 V. The concentration of dissolved ozone was monitored with a portable photometer (CHEMATEST 30, SWAN Analytische Instrumente, Hinwil, Switzerland) utilizing the DPD method (colorimetric method using *N,N*-dialkyl-1,4-phenylenediamine, ISO 73932:2017).

After applying the electrical current to the diamond electrodes, the ozone concentration increases within about 30 min to an asymptotic saturation value of 5 mg L⁻¹.

Effect of ozonated water on *P. penetrans* and cuticle-attached bacteria

The susceptibility of *P. penetrans* to various concentrations of ozone was tested. Mixed infective stages of *P. penetrans* nematodes were extracted and purified from a two-month culture on carrot discs using a Baermann funnel. The extracted nematodes were washed with 20 mL sterile tap water on a 5- μ m sieve (Cell-Trics1 filters, Sysmex, Norderstedt, Germany) and collected in sterile tap water. Ozone was electrochemically generated from ice-cold demineralized water. At various time points during ozone generation over 12 min (electrode on) or the subsequent decay of the generated ozone over 88 min (electrode off), the ozone concentration was determined and 100 μ L or 400 μ L ozonated water were added to microtubes containing 500 nematodes to reach a final volume of 500 μ L. Nematodes in demineralized water served as control. Thereby, the nematodes were exposed to ozonated water at 48 ozone concentrations between 0.03 and 4.2 mg L⁻¹. Four hours after treatment, dead and live nematodes were quantified microscopically based on their body shape (non-mobile straight- to c-shaped animals were recorded as dead nematodes, while mobile worm-shaped animals were recorded as live nematodes). To determine the number of nematode-attached bacteria that survived the ozone treatment, a volume of 100 μ L of the nematode suspension was plated on R2A media supplemented with 10 mg L⁻¹ cycloheximide. R2A allows for growth of a large variety of bacterial species (Massa et al. 1998). The R2A plates were kept at 28 °C for two days before counting the colony forming units.

Effect of ozonated water on the invasion of *P. penetrans* into soybean roots

In our second experiment, we investigated to what extent ozonated water affected the invasion of *P. penetrans* into soybean roots. Mixed infective stages of *P. penetrans* were prepared as described. Twenty pots were filled with 150 mL sand (< 1 mm) and inoculated with 10 mL water containing 2000 mixed stages of *P. penetrans*. Half of the pots were treated with 10 mL freshly produced ozonated water with $4 \pm 0.4 \text{ mg O}_3 \text{ L}^{-1}$ and the other pots with 10 mL of the untreated demineralized water as control. Pots were planted with 10 days old soybean seedlings and kept in the greenhouse at 24 °C and 16 h photoperiod. All pots were watered as needed every 2–3 days. Plants were sampled for plant growth evaluation and counting of nematodes in the roots ten days after transplanting. The whole root systems were washed and stained with 1% acid fuchsin (Bybd et al. 1983) to count the numbers of nematodes inside the roots using a SZX12 stereomicroscope at $40\times - 80\times$ magnification (Olympus, Hamburg, Germany).

Effect of ozonated water on nematodes in ARD soil

In the third experiment, we evaluated the effect of ozonated water on total nematode populations in soil, comprising plant-parasitic and free-living species. The soil was sampled from a frequently replanted orchard in the Pinneberg area (53° 41' 57.1" N 9° 40' 59.4" E) and had a high potential to induce nematode-mediated ARD (Mahnkopp et al. 2018). Since 2009, rootstocks of the cultivar 'Bittenfelder Sämling' were planted repeatedly in a two years cycle. The site had four plots randomly arranged with plants showing severe ARD symptoms. From each ARD plot, the soil was sampled around the roots of three individual plants at a depth of 0–30 cm and sieved through a 5 mm mesh to homogenize soil. This resulted in 12 replicate samples of ARD soil. Ozonated water with a concentration of $5 \pm 0.5 \text{ mg O}_3 \text{ L}^{-1}$ was prepared, and 30 ml were applied to 200 ml of ARD soil. Controls received 30 mL of demineralized water. After incubation overnight, a subsample of 100 mL soil was placed on a Baermann funnel filled with water to about 5 mm above the mesh. Nematodes that actively moved through the filter were collected over seven days. The soil was stored after ozone treatment at 4 °C to test after four weeks whether nematode populations start to recover. The low temperature was chosen to avoid microbial growth which may affect the nematodes. Active nematodes including all trophic groups were extracted by the Baermann technique and counted under a SZX12 stereomicroscope at $40\times - 80\times$ magnification (Olympus, Hamburg, Germany).

In the fourth experiment, we tested the response of nematodes in ARD soil to varying concentrations of ozonated water. Ozonated water was freshly prepared to drench 200 mL soil portions in plastic bags with 30 mL containing 24, 48, 72, 96, or 120 $\mu\text{g O}_3$ corresponding to 0.12, 0.24, 0.36, 0.48, or 0.60 mg L^{-1} soil, respectively. Controls received 30 mL of demineralized water. Soil from four replicate ARD plots was used for each treatment. After incubation overnight, subsamples of 100 mL soil were placed on a Baermann funnel filled with water to about 5 mm above the mesh, and nematodes actively moving through the filter were collected over seven days and microscopically counted. The remaining 100 mL soil samples were further incubated for four weeks at 4 °C before placing it on Baermann funnels to quantify the active nematodes.

Effect of ozonated water on mitigating apple replant disease

ARD soil was treated once or twice with ozonated water to test whether nematode-mediated growth suppression of apples plants could be alleviated. Portions of 300 mL ARD soil were drenched with 30 mL of $5 \pm 0.5 \text{ mg O}_3 \text{ L}^{-1}$, or 30 mL demineralized water as control. Half of the soil samples received a second treatment with ozonated water. The soil was transferred to pots and planted with apple rootstocks (M26, Malling 26) that have been propagated in *in vitro* culture, acclimatized, and grown to 5 cm shoot length. Pots were placed in randomized complete block design in a greenhouse and watered every 2–3 days, as required. Fertilizers NPK (+ Mg) [15:10:15(+2)] (0.5 g L^{-1}) and 36% calcium (2 g L^{-1}) were applied weekly. The greenhouse conditions were $22 \pm 2.5 \text{ °C}$, $60\% \pm 8.7\%$ relative humidity, and a 16 h photoperiod. Shoot length, shoot fresh mass, number of leaves, leaf fresh mass, and root fresh mass were determined nine weeks after planting. Overall, this experiment comprised 20 replicates for the 1 \times ozonated water treatment, 14 replicates for the 2 \times ozonated water treatment, and 17 replicates for the water control (some of the plants did not survive transplanting, so that numbers of replicates differed between treatments).

In the sixth experiment, we compared the effect of soil treatment by ozonated water and autoclaving on ARD symptoms of apple plants. For this purpose, portions of 300 mL ARD soil were filled into bags and disinfected with 30 mL $5 \pm 0.5 \text{ mg O}_3 \text{ L}^{-1}$ dissolved ozone or sterilized by autoclaving (20 min at 134 °C, 3 pulses to remove entrapped air). Water treated ARD soil served as control. The treated ARD soil was immediately planted with M26 apple rootstocks and grown as described above. Pots were sampled ten weeks after transplanting to determine shoot length, shoot fresh mass, number of leaves, leaf fresh mass, and root fresh mass. Overall, this experiment comprised 19 replicate pots each

for ozone and heat treatment, and 20 replicate pots for the untreated control.

Statistical analysis

Statistical analyses were done using the software package SAS 9.4 (SAS Institute Inc., Cary, NC, United States of America). LC_{50} concentrations and 95% fiducial limits were determined using the procedure PROBIT on Log_{10} (dose). The data significantly fitted the probit-log(dose) regression ($P < 0.0001$). The procedure GLIMMIX was used for analyses of variance by fitting data to generalized linear models (GLM). For count data as number of nematodes, colony forming units, or leaves, a Poisson distribution with a log link function was applied. Plant weights and shoot height were analysed with the assumption of a normal distribution without data transformation. For multiple comparisons, the P value was adjusted by the method of Tukey (ADJUST = TUKEY in the LSMEANS statement). To account for overdispersion of the data, degrees of freedom were approximated by the method of Kenward–Roger (DDFM = KENWARDROGER). An effect was regarded as significant at the type I error $P \leq 0.05$.

Results

Effect of ozonated water on *P. penetrans* and cuticle-attached bacteria

Exposed to increasing initial ozone concentrations, the percentage of live *P. penetrans* in suspension decreased (Fig. 1). Probit analysis revealed a LC_{50} of 0.61 mg L^{-1} ozone (95% fiducial limits: $0.602\text{--}0.624 \text{ mg L}^{-1}$). More than 99% of the nematodes were killed at ozone concentrations above 2 mg L^{-1} . The number of bacteria that was naturally attached to the cuticle of the nematodes was determined as plate counts on agar media. The bacteria were even more affected by increasing concentrations of ozone in the suspension than the nematodes (Fig. 1). Probit analysis revealed a LC_{50} of 0.27 mg L^{-1} ozone (95% fiducial limits: $0.258\text{--}0.272 \text{ mg L}^{-1}$). This suggested a significantly higher susceptibility of bacteria compared to nematodes, as expected due to direct exposure of bacterial cells in contrast to nematodes that are protected by a cuticle.

Effect of ozonated water on the invasion of *P. penetrans* into soybean roots

Pratylenchus penetrans was highly susceptible to ozone in water. However, in soil the exposure of nematodes to ozone may be reduced. We treated 150 mL soil in pots containing 2000 *P. penetrans* with 10 mL ozonated water.

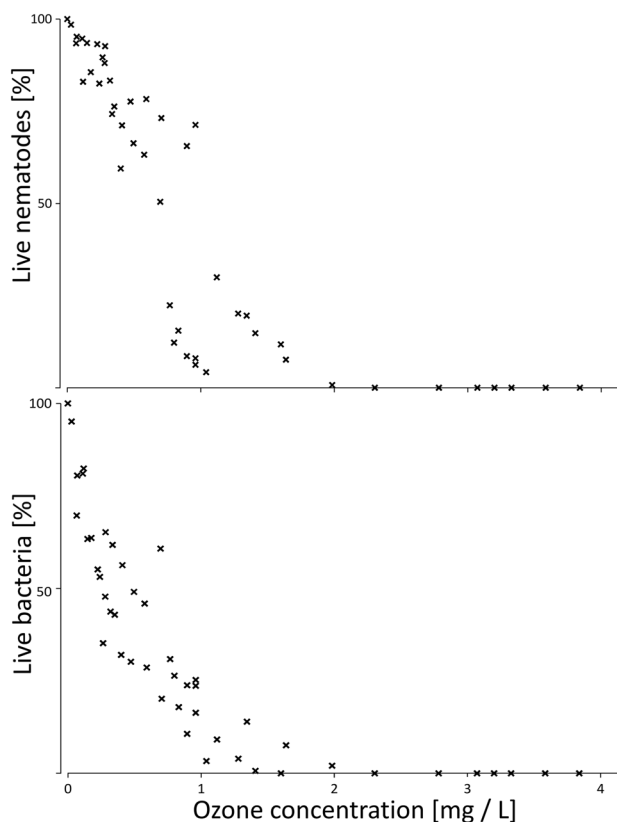


Fig. 1 Lethal doses relation of ozone towards *Pratylenchus penetrans* and its cuticle-attached bacteria in ozonated water suspension

Soybean seedlings that were planted a day after soil treatment showed significantly less nematodes that penetrated the root (Table 1). Consequently, shoot length and root weight revealed a significantly increased plant growth in ozone treated pots compared to controls (Table 1). Shoot weight showed the same trend, but the difference was not statistically significant.

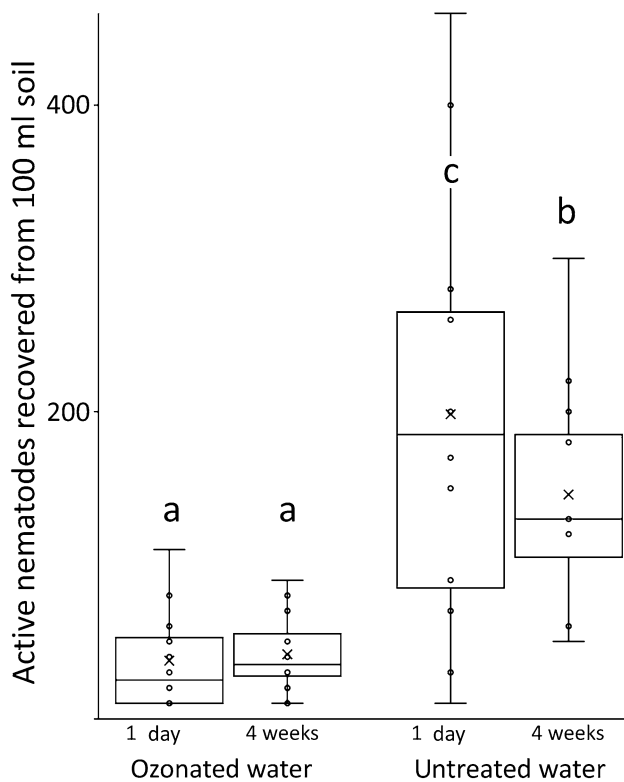
Effect of ozonated water on nematodes in ARD soil

The effect of ozonated water on total nematode populations in soil, comprising plant-parasitic and free-living species was evaluated. Drenching of 200 mL soil with 30 mL ozonated water corresponding to $0.75 \text{ mg O}_3 \text{ L}^{-1}$ soil significantly reduced the number of nematodes that actively moved through the filter on a Baermann funnel compared to the control (Fig. 2). The effect was consistent four weeks after treatment despite the increased instability of ozone in soil. The average reduction in active nematodes was 81% a week after treatment and decreased to 71% four weeks after treatment. This suggested that only few nematodes recovered from exposure to ozone within four weeks.

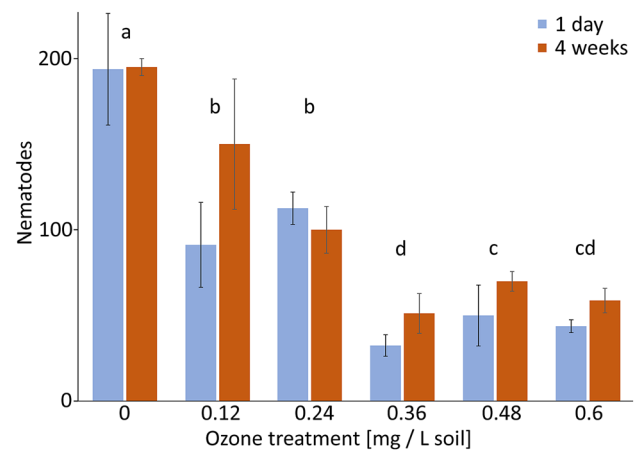
Different concentrations of ozone starting from 0.024 mg L^{-1} soil reduced the number of active nematodes in ARD

Table 1 Effect of ozone treatment of soil containing 2000 *Pratylenchus penetrans* on subsequent root invasion of nematodes and growth of soybean plants (mean \pm SE, $n = 10$)

Treatment	Plant height (cm)	Root weight (g)	Shoot weight (g)	Nematodes per root system
Ozonated water ($n = 10$)	32.7 \pm 2.1	2.3 \pm 0.1	1.7 \pm 0.2	319 \pm 33
Untreated water ($n = 9$)	26.0 \pm 2.2	1.3 \pm 0.1	1.4 \pm 0.1	791 \pm 41
GLM	$P = 0.039$	$P = 0.0001$	$P = 0.10$	$P = 0.0001$

**Fig. 2** Number of active nematodes one day or four weeks after soil treatment with ozonated water (0.75 mg ozone L⁻¹ soil), or untreated water. Different letters indicate significant differences (GLM, Poisson, log link, Tukey's adjustment, $n = 12$, $P \leq 0.05$), (—) median, (x) mean, whiskers indicate quartiles

soil, compared to the control (Fig. 3). Amounts of more than 0.6 mg ozone L⁻¹ soil did not further reduce nematode numbers. The nematode populations partially recovered within four weeks, resulting in significantly increased numbers four weeks after exposure to ozone compared to one week after treatment (GLM, Poisson, log link, factor week, $P = 0.0001$). This was most pronounced for the two lower concentrations of ozone, reaching 70% or 52% of nematode numbers relative to control soil, respectively. Treatments with 0.36–0.6 mg ozone L⁻¹ soil resulted in 26–36% active nematodes compared to the control four weeks after treatment.

**Fig. 3** Number of active nematodes in apple replant diseased soil, one day or four weeks after soil treatment with different concentrations of ozonated water, or untreated water. Different letters indicate significant differences among treatments by different amounts of ozone (GLM with treatment as factor and time as confounding factor, Tukey's adjustment, $P \leq 0.05$). Error bars indicate SE ($n = 4$)

Effect of ozonated water on mitigating apple replant disease

We tested whether the treatment of ARD soil with ozonated water improved growth of apple plants. Apple plants grown in treated soil did not show disease symptoms compared to the plants grown in untreated ARD soil. The plants showed significantly increased shoot length, shoot weight, root weight, leaf counts, and leaf weight compared to the plants grown in control soil, which showed the typical growth depression associated with apple replant disease (Table 2). The effect of the number of ozone treatments, however, was negligible, indicating that a single application substantially affects the species involved in the disease complex.

We compared the efficacy of soil treatment by ozonated water to soil sterilization by autoclaving at 134 °C in alleviating apple replant disease symptoms of plants grown in these soils. Apple plants that were grown in soil treated with ozonated water or sterilized by heat had no disease symptoms after nine weeks when compared to plants grown in untreated soil (Table 3). Plant growth was similar in ozonated water and heat-treated soils and significantly better than in untreated soil. Root weight in ozone treated soil was

Table 2 Effect of ozonated water applied one or two times to apple replant disease soil on growth of apple plantlets

Soil treatment before planting (replicate pots)	Shoot length (cm)*	Shoot fresh mass (g)	Root fresh mass (g)	Leaf count	Leaf fresh mass (g)
2×Ozonated water (14)	8.2±1.4 a	2.3±0.4 a	1.3±0.4 a	14.2±1.9 a	1.4±0.3 a
1×Ozonated water (20)	8.1±1.4 a	2.4±0.5 a	1.2±0.4 a	14.2±2.8 a	1.5±0.4 a
Untreated water (17)	6.5±0.9 b	1.3±0.4 b	0.5±0.2 b	10.6±1.8 b	0.9±0.3 b

*Mean±SD, different letters indicate significant differences (GLM with Tukey's adjustment, $P\leq 0.05$)

Table 3 Effect of ozonated water in comparison to soil autoclaving on growth of apple plantlets in soil with apple replant disease

Soil treatment before planting (replicate pots)	Shoot length (cm)*	Shoot fresh mass (g)	Root fresh mass (g)	Leaf count	Leaf fresh mass (g)
Ozonated water (19)	13.0±1.9 a	3.5±1.2 a	4.1±1.4 a	16.0±4.9 a	2.0±0.6 a
Autoclaving 134 °C (19)	13.6±2.1 a	4.5±0.8 b	4.8±1.6 a	18.3±3.1 a	2.7±0.5 b
Untreated water (20)	10.5±1.2 b	2.6±0.7 c	1.6±0.5 b	10.3±2.0 b	1.5±0.5 c

*Mean±SD, different letters indicate significant differences (Tukey's test, $P\leq 0.05$)

2.6 times higher in comparison to the untreated control. The latter showed a typical brownish discolouration. Shoot and leaf weight of apple plants grown in heat-treated soil were significantly larger compared to plants grown in soil treated by ozonated water, and this trend was also observed for shoot length, root weight, and leaf number (Table 3).

Discussion

In this study, we tested whether application of ozonated water to control nematode pests could become a promising strategy. In suspension, mixed infective stages of the plant-parasitic nematode *P. penetrans* were highly susceptible to dissolved ozone, with 50% mortality at 0.6 mg ozone L⁻¹. For juveniles of the root-knot nematode *Meloidogyne incognita*, a mortality of 91% was reported when treated twice with 0.5 mg L⁻¹ ozone dissolved in a nutrient solution for soilless cultivation of plants (Zheng et al. 2020). Eggs of *Meloidogyne enterolobii* were inactivated when suspended in 0.55 mg L⁻¹ ozone for 15 min (Landa Fernández et al. 2019). Ozone concentrations per litre soil are not directly comparable to concentrations in water, because only a minor part of the soil volume is water. Ozonated water films are unevenly distributed in soil and target organisms can be protected in cavities. Disinfection efficiency of dissolved ozone is expected to be much lower in soil than in solution, because decomposition of ozone is accelerated by oxidation of soil organic matter. In our greenhouse experiment, application of 0.27 mg dissolved ozone per litre of sand significantly affected *P. penetrans*, so that its invasion into roots of subsequently grown plants was reduced by 60%, and plant growth was significantly increased compared to controls.

Similarly, dissolved ozone was effective in growth substrate in the study of Zheng et al. (2020). Three times application of 200 ml of a nutrient solution containing 4 mg L⁻¹ ozone to eight litre of a peat-vermiculite-perlite substrate reduced juveniles of *M. incognita* by 88% and significantly affected egg hatching. Saedizadeh et al. (2019) tested the response of *Meloidogyne javanica* to ozonated water in petri dishes or in the root zone of tomato seedlings grown in pots. They reported that in both systems, ozone was efficient in controlling the nematode eggs and the second stage juveniles, leading to reduced root galling. In another study, five-month-old potato plants with a heavy infestation of root-knot nematodes were irrigated with ozonated water or non-ozonated water (Behzadi et al. 2012). After 20 days of irrigation, plants that received ozonated water developed new root systems that were no longer infected with the nematodes compared to the water irrigated plants.

We investigated whether ozonated water is efficient in treating replanted soil of tree nurseries or apple orchards to mitigate ARD symptoms of apple plants. The disease is characterized by retarded vegetative growth of apple trees and yield decline (Winkelmann et al. 2019). Plant-parasitic nematodes, especially *P. penetrans*, have been shown to affect apple plants (Jaffee et al. 1982). In a previous study, we showed that ARD is likely mediated by nematodes other than plant-parasitic nematodes (Kanfra et al. 2018). The definite cause of ARD is still not clear but multitrophic interactions of biological agents have been proposed to cause ARD (Nicola et al. 2018). Hence, designing a sustainable management measure for the disease without the known cause is difficult. In our study, growth of apple plants in replanted soil was significantly improved by a preceding treatment with ozonated water,

compared to the control. A single dose of ozonated water was nearly as effective as autoclaving of soil, while a second dose of ozonated water after a week did not further improve subsequent plant growth. Heat treatment of ARD soil at 50 °C or 100 °C was shown to alleviate the disease symptoms and improve plant growth significantly (Yim et al. 2013, 2015), but steaming of soil requires huge energy and is economically not viable (Winkelmann et al. 2019). Nutrients released from biota that were killed by ozone probably did not contribute to plant growth because the plants received sufficient fertilizer. Root systems in untreated ARD soil were reduced in volume and blackened, and the number of leaves and shoot fresh weight was decreased indicating typical growth depression associated with ARD symptoms (Grunewaldt-Stöcker et al. 2019). We showed previously that microbes attached to the bodies of specific nematodes synergistically enhanced replant disease of apple plants (Kanfra et al. 2018). Therefore, it is interesting to note that the microbes attached to the cuticle of *P. penetrans* were also affected by ozone in the present study. The number of actively moving nematodes in replanted soil was drastically reduced by doses above 0.36 mg ozone per litre of soil, which can be practically achieved. However, a fraction of 19–36% of the nematodes in soil remained active, significantly recovered within four weeks and recolonized the soil. Resilience of the soil biota after treatment with ozonated water needs further investigation. In this respect, the fast decomposition of ozone without toxic residuals is advantageous for the recovery of the soil biota. This process might be enhanced and controlled by soil amendments with composts or beneficial consortia of microbes to support the antagonistic potential of the soil against pests and pathogens (Sikora and Poca-sangre 1992).

The electrochemical generation of ozonated water by means of diamond-coated electrodes has proven to be an intrinsically safe, energy-efficient and environmentally friendly technique with high potential for agricultural applications. For generating 5 mg ozone in 1 L water, an electrical energy of only 30 Wh was consumed in this study. This excellent energy efficiency in combination with the low system requirements is a big benefit of electrochemical ozone generators based on diamond electrodes. Due to the low operating voltages, diamond-based ozone generators can be directly powered by the 12–24 V on-board voltages commonly available in agricultural vehicles. On-site generation of ozonated water, which is necessary due to the short half-life of ozone, is achieved by compact and scalable devices and can be easily controlled by adjustment of current. It offers the opportunity for replacing harmful pesticides or fumigants by a short-lived oxidant that is generated on-demand from water and cannot be overdosed. The next steps will be to adapt already existing industrial-scale diamond

ozone generators to the requirements of agricultural applications and to investigate technical and economic feasibility in outdoor operation.

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Authors' contributions Conceptualization, supervision, validation by HH and MH. Experiments done by XK, AE, HT, and SP. Writing of draft manuscript by XK and AE. Data analyses by HH, XK, and AE. Review and editing by HH and MH.

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Declarations

Conflict of interest The authors declare no competing interests.

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